© The Authors. This article is provided under the following license: Creative Commons Attribution-Noncommercial 2.0 Germany, http://creativecommons.org/licenses/by-nc/2.0/de/deed.en

Submitted: 27 February 2008, accepted: 16 March 2008, published: 27 May 2008

Cord blood - from a disposable byproduct of human birth into a precious source for life saving therapies

Gal Goldstein¹, Amos Toren¹, Arnon Nagler²

¹Pediatric Hemato-Oncology Department, The Edmond and Lily Safra children's Hospital; ²Division of Hematology and Cord Blood Bank, Chaim Sheba Medical Center, Tel Hashomer and Sackler school of Medicine, Tel Aviv University, Tel Aviv, Israel

Abstract

The review article concerns the transplantation of hematopoietic stem cells (HSCs) derived from cord blood (CB). This approach was previously used in pediatric settings. In partu procedures of CB HSCs harvesting, along with the routine methods of their quality control (i.e., HLA typing, testing for infectious pathogens) are listed in brief. Ca. 250,000 CB units are now stored in 35 blood banks in 21 countries worldwide. Some ethical problems with application of CB cells could arise during their long-term storage. The authors point to the controversies associated with the development of private cord blood banks (capacity is estimated at 600,000 CB units), due to indefinite and/or indefensible terms of their storage for eventual transplants. The specific potential of CB HSCs is limited by small sample volume; however relatively low numbers of HSCs with high proliferative activities, along with lower counts of T lymphocytes and their higher immunological tolerance enable HSC transplants at reduced rejection risk and lower GvHD rates.

Clinical experience with CB HSC transplantation is compared for different centers, where the high efficiency of this approach is shown, being, however, associated with longer terms of hematopoietic recovery when compared to bone marrow transplants. A minimal acceptable HSC CB dose is estimated as 1.5-2.5x10⁷ nucleated cells per kg body mass of a patient. The main areas of CB HSC transplantation are described, i.e., related or unrelated transplants, performed in non-cancer and malignant disorders. The authors point to scarce data comparing the efficiency of HSCs derived from cord blood versus bone marrow samples.

Special attention is paid to CB HSC transplantation in non-malignant conditions with bone marrow aplasia associated with unacceptably high non-engraftment risk. Good results of CB HSCT are demonstrated in hemoglobinopathies and mucopolysaccharidoses. When administering CB HSCs to adult patients, non-myeloablative conditioning regimens are proposed, despite the poorly defined efficiency of such an approach. An opportunity for simultaneous transplants of two or more HSC units is considered, including a unit of CB HSCs. An option of intraosseous CB HSC injection is also discussed. In vitro techniques of CB HSC expansion are under development, in spite of scarce data on their proliferative rates and differentiation ability. As an additional stimulus, injection of mesenchymal stem cells together with CB HSCs was recently proposed. In conclusion, the possible usage of normal CB HSCs to correct genetic deficiencies in children is described. CB HSCs' pluripotency may be also applied to the repair of various tissue lesions, e.g., myocardial infarction, or vascular defects.

Keywords: Cord blood, hematopoietic stem cells, harvesting, storage, transplantation, review

Introduction

The birth of every newborn human produces a precious byproduct. In the expelled placenta there is a sufficient amount of Cord Blood (CB), in which there is abundance of hematopoietic elements. In 1988 Elaine Gluckman showed that a Hematopoietic Stem Cell Transplantation (HSCT) can succeed by using CB as a source for the graft [Gluckman]. Gradually thousands of such transplantations began to be performed all over the world. Up until recently most transplantations with CB had been done in children; however the main progress in the field in the last 2 years has been achieved in adults with hematological malignancies. But even though HSCT is still the major indication for using CB, there is a growing interest in finding it as a source for non-hematopoietic stem cells (SC) for regenerative medicine, gene therapy vector, and other potential uses.

Collection of cord blood in the delivery room

CB's collection is done in the delivery room. The blood is drawn from the umbilical vein, before or after the expulsion of the placenta.

The main advantage of the collection process of CB is its simplicity. It poses no danger and causes no pain to the laboring mother or the newborn. Before the collection itself consent is given by the mother. Some centers also addend a short interview, and others use questionnaires for identification of high risk mothers [Elchalal].

Cryopreservation

After its collection CB undergoes cryopreservation. The most widely used method is the one reported by Rubinstein et al [Rubinstein]. It is based on red blood cell depletion and volume reduction. At the end of the process the total volume of each unit is 25 milliliters.

Cord blood public banks

Before freezing, CB samples undergo several tests. Every unit is screened for infectious agents, and in some banks, for relevant inherited diseases. HLA typing is usually done for A, B in serology, and DR in DNA. Some diversity exists between banks with regard to the routine tests that each unit undergoes. Additional samples are maintained in small plastic segments attached to the frozen unit in case future tests are needed.

It is estimated that today there are about 250,000 CB units frozen in 35 banks in 21 countries [Bone marrow Donors, Annual report]. CB banking is facing some challenges. The first is the scarcity of space, which is dealt with by volume reduction methods and selection of presumed optimal units, usually with higher volume. Another issue is the uncertainty regarding the period of time which units can be preserved without damage to the viability of the cells. The banks also face ethical issues. For example, if an adult disagrees with the usage of CB that his parents gave consent for donation for decades ago, what is the value of such consent? Another concern is the fate of the information that is stored in CB banks regarding donors' infectious status or the presence of genetic diseases. Is this sensitive data protected as it should be? The need for follow up is understandable, but it could also affect the diversity of ethnic pool of the donors. The ability to detect the donors might also put their families under pressure to donate more cells when the need for it might come.

The importance of public CB banking gained an official acknowledgment when the American congress decided to add \$30 million for collection of an additional 150,000 units.

Apart from the above, one of the most controversial issues is private CB banking.

Private cord blood banks

This is an ever growing trend that emerged in the early 90s. These private firms offer storage of CB units against a future need for autologous or related allogeneic transplantations. Questions have been recently being raised about whether overanxious parents are truly aware that there are no indications today for autologous cord blood transplantation (CBT). Are they informed about the slim chances for a family of ever needing a sibling CBT, and do they know about the lack of knowledge regarding the how long CB's hematopoietic SC preserve their viability while frozen?

The pace of collection in the private banks exceeds the one in the public banks. It is estimated that approximately 600,000 units are frozen privately. These facts raise questions about whether this limited source of hematopoietic SC should not be solely in public hands.

The unique properties of cord blood

Aspirates of bone marrow (BM), or the more recently used Peripheral Blood Stem Cell Collection (PBSC) product, have been traditionally used as sources for HSCT. CB has a few different qualities.

It had long been acknowledged that the more nucleated cells in a graft, the better the chances of engraftment. When taking CB into account as a hematopoietic SC source for transplantation, it is evident that it has fewer nucleated cells than other sources. Each aspirate of BM yields 750-1000 milliliters. This volume usually gives a nucleated cell dose of $2 \times 10^8 / \text{kg}$ for an average weight adult. The product of PBSC yields similar number of SC. The volume of a typical CB unit is usually only 75-150 milliliters. The nucleated cells dose is only one tenth of the BM dose, usually no more than $2 \times 10^7 / \text{kg}$, for an adult.

Another relevant component of the graft that marks the difference between CB and BM is T cells. These are considered to have a deleterious effect regarding the immune response of the graft against the recipient. The total dose of T cells (CD3+ cells) in CB units is less than a fifth of the amount in BM grafts. When comparing it to mobilized peripheral blood grafts, CB units have less than 2% of T cell dose. But while less hematopoietic SC in CB is a setback regarding HSCT, the scarceness of T cells is an advantage, with respect to the risk of graft versus host disease (GVHD) [Bensienger, Bittencourt].

The low number of SC in CB graft is masked by an excellent proliferative response. When these cells are in a dormant state and cytokines are introduced into their environment, they expand much better than hematopoietic SC of BM. This trait enables CB to produce full hematopoietic recovery of BM in myeloablated recipients [Mayani, Lewis].

The naïve nature of the immune system's cells in CB is a different issue.

The lymphocytes in CB grafts have a more tolerant nature [Nitsche, Risdon_{a,b} Roncarolo, Cohen, Garderet]. Other components of the system, such as dendritic and Natural Killer cells also have different properties, when compared to BM or adult peripheral blood [Leung, Kedereit, Sorg, Canque, Liu, Dalle]. Because of this, CB allows greater human leukocyte antigen (HLA) disparities in transplantations, with less rejection and lower rates of GVHD.

Cord blood transplantation, clinical experience

Reports on a series of CBT started to appear at the beginning of the end of the 1990s and at the beginning of the third millennium. These were based mostly on the American and the European registries, with some reports from Japanese and other institutes. Table 1 & 2 summarizes the largest clinical trials of CBT using unrelated donors [Rubinstein, Gluckman, Wagner, Michel,

Locatelli_a, Laughlin_{a,b}, Rocha, Takahashi_a, Long]. A few important concepts could be built upon results from these works. First was the notion that CB, with its limited nucleated cells dose, can produce full hematopoietic reconstitution after myeloablative conditioning. Secondly, the median time of myeloid recovery in CBT ranged from 22 to 33 days. This is a far longer period than the time in bone marrow transplantation (BMT) experience. When BM aplasia is prolonged, morbidity and mortality rates rise. The third notion was that despite the existence of a significant proportion of HLA disparity between donors and recipients, rejection and GVHD rates were surprisingly low.

So these trials proved that CB is a legitimate source for HSCT, with problematic engraftment kinetics, but less restriction to HLA matching when compared to BM.

Since each placenta contains a limited volume of blood, it follows that there is also a limited amount of nucleated cells per unit. The correlation between nucleated cell dose and transplantation outcomes was evaluated. A positive impact of cell dose on time to engraftment, and hence the overall survival, has been demonstrated in both pediatric and adult series. It is probably agreed that the minimal acceptable threshold of nucleated cells dose should be 1.5×10^7 nucleated cells/kg, but an association between dose of 3.7×10^7 nucleated cells/kg and more and faster time to neutrophil engraftment was suggested by the Eurocord [Gluckman_b, Arcese]. The New York Blood Group reported that 2.5×10^7 nucleated cells/kg is the minimal threshold for transplantation [Rubinstein].

Historically CD34+ cells counts were not part of the tests done routinely on CB units. But it is reasonable to assume that it might be so in the near future. Counting nucleated cells involves many cells that do not contribute to the engraftment potential. And indeed, Wagner et al has shown a correlation between CD34+ dose of 1.7x10⁵cell/kg and higher to rapid neutrophil engraftment and probability of engraftment [Wagner].

Related donor transplants

Although the first CBT was done from a sibling donor, related donors transplants are used less frequently in this setup. For the cure of malignant diseases CB from a sibling could be used if there is a perfect timing of a birth in the family, or a if a CB unit had been cryopreserved earlier, either by chance or by intention. In non-malignant disease there is usually more time. Families that are aware of CB as a source for transplantation might act on time when births are due.

Several reports of large series of trials have been published. These series have demonstrated that CB is a valid therapeutic option as a source for pediatric transplantation for malignant and non-malignant diseases. The probability for survival at 1 year was 0.63 (95% CI: 0.57-0.69) in the Eurocord study, and 0.61 (95% CI: 0.81-0.49) at 2 years in the ICBTR study [Rubinstein, Gluckman].

The largest of the series is a joint European and American work that compared 113 related donor CBT in children with 2052 cases of related donor BMT. Neutrophils engraftment in the CB group occurred at a median time of 26 days, compared to 18 in the BM group. Probability of myeloid recovery at day 60 was 0.89 and 0.98 in the CB and BM respectively. Children who received CB had a significantly lower risk of both AGVHD and CGVHD than those who were transplanted from BM (relative risk 0.41; p=0.001 and relative risk 0.35; p=0.02, respectively). Overall survival at 3 years was 0.64 for the CB and 0.66 for the BM group. This study demonstrated the role of related donor CBT for malignant diseases in children [Rocha_b]. Related donor CBT for non malignant diseases will be discussed in the non malignant section.

Comparison to bone marrow

No randomized trials had been conducted to compare CB with

Table 1. Umbilical cord blood transplantations - children

EFS %	os	TRM % (y)	Rel. % (y)	AGVHD (III-IV)/ Chronic %	Hematopoietic recovery			HLA loci disparity, %				Median cell dose	Median age		D. C
(y)	(y)				Neu rec. %	Media ANC PLT >:	≥500/	HL/	A loci d	ısparı	ty, %	NC x10 ⁷ /kg/ CD34+ x10 ⁵ /kg	in years	n	Ref.
EFS, (OS-NA) Gen. 48		56.8	56.0	24/21	02.7	23/75	MU	≥3	2	1	0	≥10=15% 5-9.9=22%	<2=20% 2-5=22%	861	Dukingtoir
M Aq. 2	27	(1)	56.8	24/31	92.7	28/94	MM	7	48	39	6	2.5-4.9=34% <2.5=29%/ NA	6-11=23% 12-18=14% >18=21%	801	Rubinstein
AA 21 Gen. 51 M 36		NA		39/NA	82	29/ NA		83			17	5.6 (0.8-60)/ 1.9 (0.6-78)	5.0 (0.2-15)	291	Gluckman _b
NA	47 (2)	30 (1)	37 (2)	11/9	88	23/86		2	41	43	14	2.8 (0.4-39.1)/ 3.1 (0.7-57)	7.4 (0.2-57)	102	Wagner
42 (2)	45 (2)	20 (2)	29 (2)	35**/15	78	NA		13	33	46	8	4.4 (0.4-36)/ 1.4 (NA)	6.0 (0.3-16)	95	Michel
30	NA	52	40	23/12	79	33/85*		30	NA	52	40	5.0 (1.5-46.5)/ 1.8 (0.1-78)	5.5 (1.7-14)	60	Locatelli _a

Table 2. Umbilical cord blood transplantations - adults

EFS % (y)	OS % (y)	TRM % (y)	Rel. % (y)	AGVHD (III-IV)/ Chronic %	Hematopoietic recovery			HLA loci disparity,%				Median cell dose		Median	Pts	
					Neu rec.	Mediar PLT ≥	n day	≥3	2	1	0	CD34+ x10 ⁵ /kg	NC x10 ⁷ /kg	age in years	#	Ref.
					/0	50.000	≥500									
20 (3.3)	63 (5)	22 (3.3)	40/ 50	17 (5)	~65	60	27	0	77	23	0	NA	2.2 (1-6.5)	NA	150	Laughlin _a
21 (1)	54	27 (1)	38/ NA	NA	81	NA	32		94		6	0.8 (0.01-8.9)	2.2 (1.2-7.3)	26 (15-53)	108	Gluckman _b
33 (2)	44 (2)	36 (2)	26/ 30	23	75	NA	26	4	39	51	6	NA	2.3 (0.9-6)	24.5 (15-55)	98	Rocha
26 (3.3)	51	28 (3.3)	20/ 36	5.9 (1)	90	99	27	17	54	26	3	1.2 (0.2-16.7)	2.1 (1-6.3)	31 (18-58)	68	Laughlin _b
74 (2)	9 (1)	NA	7/ 77	16 (2)	88	48	22	25	54	21	0	0.9 (0.2-9.0)	2.47 (1.1-5.29)	36 (16-53)	68	Takahashi _a
15 (3)	50	19 (3)	16/ 32	15.7	71	84	26	5	77	14	4	1.37 (0.02-12.4)	2.12 (1.1-4.4)	31 (18-58)	57	Long

BM grafts. Few retrospective reports have been published. As for children, it was shown by Eapen that 503 cases of matched CBT had better 5 DFS than 116 matched unrelated donor (8/8) BMT. Even the 5/6 matched CBT had comparable results with the BM group. An important factor was the cell dose. The group that received more than 3×10^7 nucleated cell/kg had better DFS and OS [Eapen]. It was Rocha and Gluckman who assessed leukemia-free survival at 5 years after CBT or BMT in children. 503 children received CB-either matched or mismatched. The outcome of these transplantations was compared to BMT of 282 children. Allele-matched bone-marrow transplants had similar outcomes to transplants of umbilical cord blood mismatched for either one or two antigens. Higher survival rates were demonstrated after transplants of HLA-matched umbilical cord blood [Rocha].

Recent publications have managed to evoke hopes that even in adults CBT (matched, or 1-2 HLA antigens mismatched) is as good as matched unrelated donor BMT. The reports of Laughlin, Rocha and Takahashi in late 2004 compared a large series of adult patients who received unrelated CB or BM. Outcomes of CBT were similar, and in certain aspects superior, to unrelated donor mismatched BMT.

Laughlin found that patients receiving mismatched CB had similar treatment-related mortality, treatment failure, relapse and overall mortality rates, to those received mismatched BM. Rocha compared matched unrelated donor of BM with CB. He found no differences in treatment-related mortality rates, relapse and leukemia-free survival rates between them. These results may refine the accepted approach for unrelated donor search. Many believe that a search for a BM donor and a CB unit should generally be started simultaneously and CB (matched or mismatched in up to 2 HLA antigens) should be preferred if matched BM donor can not be found within a reasonable period of time [Rocha, Laughlin, Takahashi]. In late 2006 Takahashi et al published the first report

of adult transplantation with CB as a first option for non related donor graft. The Japanese group transplanted 100 adults with hematological malignancies with CB, if they had no matched related donor. Results of the CBT were compared to matched related BM or peripheral SC transplantations. The outcome was similar in all groups. Whether this interesting approach is feasible in all cases of patients with no matched related donor, relies upon further reports from other ethnic groups [Takahashi,].

CBT for non malignant diseases

HSCT can offer the only true chance for cure in many nonmalignant diseases. CB offers some unique advantages in the area of transplantations for non malignant diseases. Many of these patients are children. This makes nucleated cells doses satisfactory in most of the cases. Moreover, rareness of GVHD tempts the preference of CB, especially in an unrelated donor setup. As opposed to HSCT for malignant disease, there is no presumed benefit from the Graft Versus Leukemia effect of GVHD. On the other hand, CB is a less attractive option for transplantation for bone marrow failure syndromes. There are high rates of graft rejection in HSCT in these diseases. When adding the negative impact of CB's tendency for delayed engraftment, it is regarded by some as a problematic solution for such patients. This was demonstrated in the work of Rocha et al. In a related donor setting, and definitely with unrelated donors, for bone marrow failure syndrome patients, it was clear that engraftment, and therefore event free survival (EFS) rates are not acceptable. The probability of myeloid engraftment at day 60 was not more than 67% for patients that were given related donor grafts, and it was 36% in unrelated donor-CBT. Only 33% of the Fanconi anemia patients engrafted [Aker]. Better results were reported by the European group when they summarized unrelated CBT for Fanconi patients. Although only 12 of the 93 cases were HLA identical; 60% of the patients engrafted by day 60. A positive impact of Fludarabine based regimens, cell dose, and CMV negative recipients was seen [Gluckman_d].

Some limited experience was gained by us with a few bone marrow failure syndromes, namely Fanconi anemia. We observed high rates of event free survival (EFS), especially in children who received a matched family donor transplant [Goldstein].

In one case we used a novel strategy of pre-implantation genetic diagnosis for one of the patients. This method, which is based on CBT, could pave the way for many malignant and non-malignant diseases [Bielorai].

Although the role of HSCT for Thalassemia in the era of newer iron chelating agents is yet to be determined, this strategy is still being practiced widely in an attempt to cure this hemoglobinopathy. Locatelli et al reported results of related CBT in 44 children with hemoglobinopathies (Thalassemia and Sickle Cell Disease), and showed that this procedure is feasible. High rates of engraftment (89% at day 60) and EFS (79% for Thalassemia and 90% for Sickle Cell Disease) were achieved [Locatelli,].

As for CBT in inborn errors of metabolism, Staba et al reported impressive results in children with Hurler syndrome who were given unrelated donors CB grafts. Even though 19 of the 20 patients received mismatched grafts, high rates of engraftment were reported (at 2.4 years follow-up, 85%). This was probably due to the relatively high nucleated cells doses (median of 10.5×10^7 nucleated cells/kg). The disease itself was cured, as could be seen in all 17 patients who were alive, and had normal peripheral-blood α -L-Iduronidase activity [Staba]. Recently a report of a case of a child who was cured of Wolman disease by a CBT was published [Stein].

CBT for the cure of Sickle Cell Anemia was reported recently by a French group. Importantly the authors noticed that after a 6 year follow up the group of patients that received a CB graft did not develop the main contributing factor for the morbidity, GVHD [Bernaudin].

Investigational approaches in cord blood

Most patients needing HSCT are adults. For these heavier patients CB is a problematic solution because of the relatively low cell dose. Various strategies are being attempted in order to lower the toxicity of the conditioning regimen. This could be achieved either by lowering its intensity, or by hastening engraftment.

Reduced intensity conditioning

The practice of HSCT with reduced intensity conditioning (RIC) has emerged in the adult population. These older patients usually have pre-existing morbidities.

By reducing the intensity of the preparative regimen it has been shown that treatment-related morbidity and mortality rates could be lowered. The concept behind this is based on the assumption that in certain cases the immunological impact of the graft is more important than the ablative power of the conditioning regimen.

Experience with transplantations using RIC, though follow up

time is still short, have shown encouraging results. Patients who benefit the most from RIC are those with diseases of a more indolent nature.

Few studies of RIC-CBT in adult and pediatric patients have been published. The major conclusion that could be drawn from these series is that RIC is feasible in CBT. Graft rejection happened mainly in cases in which the accumulative chemotherapy dose experienced by the recipients prior to the transplantation itself was low. Though survival rates are low, it must be emphasized that most studies included mainly high risk, heavily treated patients. GVHD rates correlated with unrelated donor BMT. Another encouraging finding is the lower than expected rates of treatmentrelated mortality at 100 days post-transplantation. Because of the small number of patients, and diversity of methods, conclusions regarding the optimal RIC conditioning regimen, or the GVHD prophylaxis, can not be drawn at this point. Even if it is definitely too early to recruit patients for RIC-CBT outside clinical trials for selected patients, these protocols could offer an alternative for selected patients [Miyakoshi, Del Toro, Chao, Bradley, Barker, Ballen, Yuji].

Engraftment hastening

The idea of shortening the period from transplantation to myeloid recovery is at the basis of many strategies. Some have shown preliminary encouraging results in the laboratory, in animal models, and even in clinical trials.

Transplantations with double cord blood units

Many recipients receive more than one partially matched CB units where the cell dose in each is not sufficient. In many cases the sum of these units provides an adequate number of SC. It has been shown in animal models that two CB units provide high rates of engraftment [Nauta]. Some studies have used this strategy for high risk adult patients who received two mismatched CB units. Many believe that this strategy could pave the way for lowering treatment-related mortality rates in CBT. In most of these trials two encouraging facts were observed: stable mixed chimerism, and no mutual rejection of mismatched units [Barker, De Lima, Gryn, Ballen,]. Brunstein et al have shown that by using a nonmyeloablative regimen for CBT in adults, the OS of the group that received 2 units was higher than the patients who received 1 unit. In this study 92% of the patients achieved neutrophil recovery, at a median time of 12 days [Brunstein]. Interestingly, sustained hematopoiesis after double CBT is usually derived from a single donor. The relative percent viability, the infused number of NC and CD34⁺ cell doses, and the donor–recipient HLA-disparity are not helpful in predicting which of the two CB units will predominate. Although early data suggested that the dominant unit had a higher median infused CD3+ cell dose, this observation has not persisted with investigation of a larger cohort of patients. Order of infusion, location of HLA mismatch, ABO blood group and/or sex mismatch also did not have a predictive effect on engraftment.

Double CBT can potentially produce a better graft versus leukemia effect. This was demonstrated in a study of the University of Minnesota. They compared leukemia patients who received 2 units of CB to those who received a single unit. The group who received the double CBT had a lower risk for relapse. It is still

not known if the relatively high degree of HLA mismatch in this setting is responsible. It might also be a consequence of non-HLA disparity, such as KIR mismatch, between the CB units and the recipient, or between themselves [Verneris].

Double unit transplantation has become a major breakthrough in the field of CB during the last 2 years. Several 2 arms protocols for using double units are on their way. Whether these expectations are justified depends on preliminary results of these trials.

Co transplantation with a Haploidentical donor

Relaying on the assumption that almost every patient has a donor, namely a parent that has a similar HLA type of one of his haplotypes, Magro et al have succeeded in transplanting CB together with a Haploidentical graft. They succeeded in inducing a rapid engraftment via a BM transplant. By administering only a small dose of Haploidentical SC, the Spanish team managed to induce a temporary engraftment only. These cells were rejected later, due to their low dose and significant HLA disparity, allowing engraftment of the CB graft. 69% of these high risk patients survived at 4 years [Magro].

Intra-osseous transplantation

One of the obstacles to a short period of engraftment is the possibility that the homing process is influenced by anatomical barriers. It has been suggested that intra BM injection of the graft could induce a rapid engraftment. This has been shown to be feasible, and has improved engraftment kinetics in BMT in adults [Hagglund]. Time will tell if intra osseous transplantation could shorten the way for CB's SC into the BM, and therefore improve time to engraftment, as has been shown in animal models [Yahata, Wang].

Ex vivo expansion of hematopoietic stem cells

In vitro studies had shown that SC proliferate with the addition of cytokines. But uncontrolled expansion is not biologically satisfactory, since maturation and differentiation of SC occur in these conditions. The SC proliferate and become committed to specific hematopoietic cell lines. Such cells lack what is known as "long term population ability." The optimal composition of the cytokine-rich media of the ex vivo expansion process is an important challenge for researchers to face. It has been demonstrated by Shpall et al that co transplantation of ex vivo expanded and non-manipulated grafts are feasible. But in spite of this, improvements in engraftment kinetics, are yet to be achieved [Bruno, Piacibello, Kohler, Shpall_{ab} Jaroscak, Pecora].

Different attitudes have been taken in order to refine the expansion process, namely: co-culturing with different cells as feeder layers [Zhang, Chute], selection of SC for the expansion [Forraz], and multiplying the proliferative potential by performing a two step harvesting technique [McNiece_a, Pick]. None of these strategies have yet been introduced into clinical trials.

A somewhat more promising field is interference with the differentiation of expanded SC. Reports have been published recently regarding ex vivo expansion with copper chelator, Tetraethylenepentamine (TEPA), which enhances the long term populating ability of the CB cells. Following large scale

experiments, this appealing approach has been introduced into the clinic in phase I trials. Preliminary encouraging results of this trial with no significant toxicity were presented recently [Peled_{a,b,c}, Grynspan, Shpall c]. A Phase II multi center study has just started and the first 3 adult patients with hematological malignancies have already been recruited (De Limab). The same concept was behind the experiment held by Nolta et al, when they co-cultured primitive CB's SC (CD34+ CD38-) together with a feeder layer of immortalized murine stromal cell-line AFT024. This method has yielded high rates of myeloid and lymphoid engraftment in a NOD/SCID mouse xenograft model [Nolta]. Other molecules that play major roles in the differentiation of hematopoietic cells, and might be used in the future for ex vivo expansion of CB are Gfi-1 and some of the Notch ligand protein family [Laureta Hock].

Novel methods have been studied recently with the use of epigenetic factors. Silencing of genes could be a consequence of methylation of their promoters or deacetylation of histones. By trying to inhibit these processes, reactivation of some genes could augment the hematopoietic SC's self renewal potential. Recent publications have shown some success in the in vitro repopulating potential of CB when using histone deacetylase inhibitors, such as Valproic acid [Milhem, De Felice]. This strategy is the basis of a clinical study which has recruited the first patients (personal communications).

Cord Blood, Umbilical cord, and Mesenchymal Stem Cells

As their unique qualities are revealed, the interest in mesenchymal stem cells (MSCs) is growing continuously. These cells are nonhematopoietic stromal cells that are capable of differentiating into, and contribute to the regeneration of, mesenchymal tissues, but possibly also to other tissue lineages. They have an in vitro expansion ability while their growth and differentiation potential is maintained. Currently it is expected that they could be used for tissue repair and regenerative medicine. MSCs have shown that they can modulate immune response both in vitro and in vivo. Preliminary studies are on their way for using MSCs as an anti GVHD prophylaxis. It was doubted that these cells could also play a roll in treating GVHD. It has also been postulated that these cells could be used for other immune mediated diseases. MSCs are used as a growing medium for ex vivo expansion of other cells [McNiece, Robinson]. Le Blanc et al showed that MSCs could be transfused in parallel to HSC grafts and demonstrated fast engraftment [Le Blanc]. Finally, MSCs are considered to be candidates as a vector for gene therapy.

Until recently only BM and adipose tissue were considered as a source for MSC. In the last few years it had been shown that CB contains MSC [Lee]. MSC from other sources has been demonstrated to have suppressive effect on T cells [Li]. Few studies have focused on the different properties of MSC originating from CB. Their tendency to differentiate into specific tissue, their genomic expression, and proliferative response, are all different from BM or adipose tissue MSC [Chang, Kern].

When considering the expulsion of the placenta at the end of delivery as a waste of a precious source of SCs, it is not only the CB itself that should be regarded as such. The Wharton jelly in the umbilical cord has also been found to be a source for MSCs [Friedman, Secco].

Cord blood uses in other fields

Gene transfer is an exciting new field in which CB could serve as a vector for correcting inborn genetic errors, or replace infected hematopoietic SC, such as in the case of HIV. Its availability, proliferative response, and engraftment potential, make it an appealing candidate for these uses [Ikeda]. Clinical trials of gene transfer to Adenosine Deaminase deficient Severe Combined Immunodeficiency children relied on BM and CB as a hematopoietic SC source. This method faced some obstacles that continue to prevent it from curing these patients [Kohn, Aiuti].

Another—to date only investigational—field is the potential non-hematopoietic use of hematopoietic SC. In recent years much interest has been focused on the ability of hematopoietic SC to differentiate into (or as some claim, to fuse with) cells of other tissues. It was demonstrated that cells with pluripotent differentiation potential could be found in CB [Pesce, Newsome, Goodwin]. CB has been suggested to have a role in improving performance of rats who were subjected to brain infarct [Vendrame]. CB is also considered by some to be a source of SC for regeneration of ischemic heart tissue by differentiation processes or neoangiogenesis [Ma]. It is too early to define whether SC's plasticity might have clinical benefits in repairing injured tissues, but this application is at the center of great interest and controversy.

Discussion

CB has become a legitimate source, not only for HSC for transplantations, but also for other uses. The experience gained in the last twenty years of work with CBT has shown us its advantages, as well as its setbacks.

Unlike BM donations, CB's collection is easier and poses no danger to the donor. In CB banks there is a greater proportion of ethnic minorities than in BM registries. It also has greater availability in an unrelated donor HSCT due to its shorter donor search time. Lesser risk for transmission of infectious agents in the transplantation process is another benefit of CB. There is no doubt that fewer HLA restrictions in unrelated transplantations is its main advantage. This fact allows successful transplantations with acceptable rates of graft failure and GVHD.

On the other hand, there is a slim potential for disease transmission, namely genetic, in CBT setup. In CBT there is almost no option for a second transplantation, or any boost of cells. A troubling disadvantage of CB is its low number of hematopoietic SC in each unit. This has proven to be a crucial point that has a direct relationship to relatively high rates of treatment-related mortality rates in CBT. This point is further emphasized within the setting of adult transplantations.

From the data collected in several series of CBT for both malignant and non malignant diseases it appears that CB can be used as a SC source in several settings.

The most urgent need for SC is transplantation for malignant diseases from unrelated donors. It is an acceptable approach to search first for BM donor. If a 5/6 or better HLA match can not be found, or progressive disease status does not allow completion

of the search, then a CBT of 4/6 HLA match or better should be performed. This of course depends on a minimal cell dose of $2x10^7$ nucleated cells/kg per CB unit. Cell dose has greater relevancy in adult transplantation setup. Skepticism about the possibility of CBT in heavier patients might fade as newer strategies could overcome SC scarceness of nucleated cells in CB. At this stage the most appealing strategy is the double unit CBT. By receiving 2 CB units many adults could be transplanted with a reasonable time to engraftment. Time will tell if other methods could offer a solution for a better outcome in CBT for adults.

Impressive progress is constantly being achieved in the field of CBT. CB is still considered a second best choice for HSCT, but as newer reports are being published it is not so obvious whether it could not be preferred over BM. Interesting data in children showed that a perfect match (6/6) of CB could be the best choice. If larger studies can confirm this, we might see CB becoming the first option for transplantation in certain conditions.

For non malignant disease CB is a very good option, especially for the smaller patients. Caution should be practiced when using CB for bone marrow failure syndrome, though again it seems that larger units and better preparative regimens may overcome the tendency for graft failure.

Future uses of CB may not be just for HSCT. Time will tell if the fields of gene therapy and non hematopoietic injured tissues repair also benefit from the use of CB cells.

References

Aker M, Varadi G, Slavin S, Nagler A. Fludarabine-based protocol for human umbilical cord blood transplantation in children with Fanconi anemia. J Pediatr Hematol Oncol. 1999;21:237-239.

Aiuti A, Slavin S, Aker M, Ficara F, Deola S et al. Correction of ADA-SCID by stem cell gene therapy combined with nonmyeloablative conditioning. Science 2002;296:2410-2413.

Arcese W, Rocha V, Labopin M, Sanz G, Iori AP et al. Eurocord-Netcord Transplant group Unrelated cord blood transplants in adults with hematologic malignancies. Haematologica 2006 Feb;91(2):223-30.

^a Ballen KK, Becker PS, Emmons RV, Fitzgerald TJ, Hsieh CC, et al. Low-dose total body irradiation followed by allogeneic lymphocyte infusion may induce remission in patients with refractory hematologic malignancy. Blood 2002;100:442-450.

_b Ballen KK, Spitzer TR, Yeap BY, McAfee S, Dey BR et al. Double unrelated reduced-intensity umbilical cord blood transplantation in adults. Biol Blood Marrow Transplant. 2007 Jan;13(1):82-9.

Barker JN, Weisdorf DJ, DeFor TE, Blazar BR, Miller JS et al. Rapid and complete donor chimerism in adult recipients of unrelated donor umbilical cord transplantation after reduced intensity conditioning. Blood 2003;102:1915-1919.

_b Barker JN, Weisdorf DJ, Wagner JE. Creation of a double chimaera after transplantation of umbilical-cord blood from two partially matched unrelated donors. N Engl J Med. 2001:344.1870-1871.

^c Barker JN, Weisdorf DJ, DeFor TE, Davies S, Verfaillie CM et al. Impact of multiple unit unrelated donor umbilical cord transplantation in adults: preliminary analysis of safety and efficacy. Blood 2001;98 (suppl 1):2791(abstract).

Bensienger WI, Martin PJ, Storer B, Clift R, Forman SJ, et al.

Transplantation of bone marrow as compared with peripheral-blood cells from HLA-identical relatives in patients with hematologic cancers. N. Eng. J. Med. 2001,344,175-181.

Bernaudin F, Socie G, Kuentz M, Chevret S, Duval M et al. Long-term results of related myeloablative stem-cell transplantation to cure sickle cell disease. Blood 2007 Oct 1;110(7):2749-56.

Bielorai B, Hughes MR, Auerbach AD, Nagler A, Loewenthal R et al. Successful umbilical cord blood transplantation for Fanconi Anemia using preimplantation genetic diagnosis for HLA-matched donor. Am J Hematol. 2004;77:397-399.

Bittencourt H, Rocha V, Chevret S, Socie G, Esperou H et al. Association of CD34 cell dose with hematopoietic recovery, infections, and other outcomes after HLA-identical sibling bone marrow transplantation. Blood 2002,99,2726-2733.

Bradley MB, Satwani P, Baldinger L, Morris E, van de Ven C et al. Reduced intensity allogeneic umbilical cord blood transplantation in children and adolescent recipients with malignant and non-malignant diseases. Bone Marrow Transplant. 2007 Oct;40(7):621-31.

Bone marrow Donors, Annual report, 2003.

Bruno S, Gammaitoni L, Gunetti M, Sanavio F, Fagioli F et al. Different growth factor requirements for the ex vivo amplification of transplantable human cord blood cells in a NOD/SCID mouse model. J Biol Regul Homecost Agents. 2001;15:38-48.

Brunstein CG, Barker JN, Weisdorf DJ, DeFor TE, Miller JS et al. Umbilical cord blood transplantation after nonmyeloablative conditioning: impact on transplantation outcomes in 110 adults with hematologic disease. Blood 2007 Oct 15;110(8):3064-70.

Canque B, Camus S, Dalloul A, Kahn, E. Yagello, M et al. Characteristic of dendritic cell differentiation pathway from cord blood CD34(+)CD7(+)CD45(+) hematopoietic progenitors cells. Blood 2000;96:3748-3756.

Chang YJ, Shih DT, Tseng CP, Hsieh TB, Lee DC et al. Disparate mesenchyme-lineage tendencies in mesenchymal stem cells from human bonemarrow and umbilical cord blood. Stem Cells 2006 Mar;24(3):679-

Chao NJ, Koh LP, Long GD, Gasparetto C, Horwitz M et al. Adult recipients of umbilical cord transplants after nonmyeloablative preparative regimens. Biol Blood Marrow Trans. 2004;10:569-575.

Chao PA, Bearman SI, Jones R, Nieto Y, Cagnoni PJ et al. Nonmyeloablative hematopoeitic cell transplant using cord blood. Blood 2001;98(suppl):2794abstract.

Chute JP, Muramoto G, Fung J, Oxford C. Quantitative analysis demonstrates expansion of SCID-repopulating cells and increased engraftment capacity in human brain endothelial cells. Science 2004;22:202-215.

Cohen SB, Madrigal JA. Immunological and functional differences between cord and peripheral blood. Bone marrow Transplant. 1998;2(Suppl 3):S9-S12.

Dalle JH, Menezes J, Wagner E, Blaqdone M, Champaqne J et al. Characterization of cord blood natural killer cells: Implication for transplantation and neonatal infetions. Pediatr Res. 2005;57:649-655.

De Felice L, Tatarelli C, Mascolo MG, Gregorj C, Agostini F et al. Histone deacetylase inhibitor valporic acid enhances the cytokineinduced expansion of human hematopoietic stem cells. Cancer Res. 2005;65:1505-1513.

De Lima M, St John LS, Wieder ED, Lee MS, McMannis J et al. Double-chimaerism after transplantation of two human leucocyte antigen mismatched, unrelated cord blood units. Br J Haem. 2002;119:773-776.

De Lima M, McMannis J, Gee A, Komanduri K, Couriel D et al. Transplantation of ex vivo expanded cord blood cells using the copper chelator tetraethylenepentamine: a phase I/II clinical trial. Bone Marrow Transplant. 2008 Jan 21 [Epub ahead of print].

Del Toro G, Satwani P, Harrison L, Cheung YK, Brigid D, et al. Pilot study of reduced intensity conditioning and allogeneic stem cell transplantation from unrelated cord blood and matched family donors in children and adolescellsent recipients. Bone Marrow Transplant. 2004;33:613-622.

Eapen M, Rubinstein P, Zhang MJ, Stevens C, Kurtzberg J, et al. Outcomes of transplantation of unrelated donor umbilical cord blood and bone marrow in children with acute leukaemia: a comparison study. Lancet 2007 Jun;369(9577):1947-54.

Elchalal U, Fasouliotis SJ, Shtockheim D, Brautbar, C. Schenker, JG et al. Postpartum umbilical cord blood collection for transplantation: a comparison of three methods. Am J Obstet Gynecol. 2000;182:227-232.

Forraz N, Pettengell R, McGuckin CP. Characterization of a lineagenegative stem-progenitor cell population optimized for ex vivo expansion and enriched for LTC-IC. Stem Cells 2004;22:100-108.

Friedman R, Betancur M, Boissel L, Tuncer H, Cetrulo C et al. Umbilical cord mesenchymal stem cells: adjuvants for human cell Transplantation. Biol Blood Marrow Transplant. 2007 Dec;13(12):1477-8.

Garderet L, Dulphy N, Douay C, Chalumeau, N.; Schaeffer, V et al. The umbilical Cord blood alphabeta T-cell repertoire: characteristics of a polyclonal and naïve but completely formed repertoire. Blood 1998;91:340-346.

Gluckman E, Broxmeyer HA, Auerbach AD. Hematopoietic reconstitution in a patient with Fanconi's anemia by means of umbilicalcord blood from an HLA-identical sibling. N Engl J Med. 1989;321:1174-

Gluckman E, Rocha V, Chevret S. Results of unrelated umbilical cord blood heamatopoeitic transplantation. Rev Clin Exp Hematol. 2002;5:87-

Gluckman E, Rocha V, Boyer-Chammard A, Locatelli F, Arcese W, et al. Outcome of cord-blood transplantation from related and unrelated donors. Eurocord Transplant Group and the European Blood and Marrow Transplantation Group. N Engl J Med. 1997;337:373-381.

Gluckman E, Rocha V, Ionescu I, Bierings M, Harris RE, et al; Eurocord-Netcord and EBMT. Results of unrelated cord blood transplant in fanconi anemia patients: risk factor analysis for engraftment and survival. Biol Blood Marrow Transplant. 2007 Sep;13(9):1073-82.

Goldstein G, Bielorai B Nagler A, Hutt D, Neuman Y, et al. Cord blood transplantation in children – a single institute experience. Bone marrow Transplant. 2005;35(suppl 2):S366.

Goodwin HS, Bicknese AR, Chien SN, Bogucki BD, Quinn CO et al.Multilineage differentiation activity by cells isolated from umbilical cord blood: expression of bone, fat, and neural markers. Biol Blood Marrow Transplant. 2001;7:581-588.

Gryn J, Harris DT, Shadduck RK, Raymond J, Ziegler Z et al. Multiple unmatched umbilical cord units (MUCs) for adult allogeneic transplantation. Blood 98(suppl);2001:2792 (abstract).

Grynspan L, Peled T, Rosenheimer-Goudsmid N, L. Hana, N. Hasson, J. et al. Cord Blood Derived CD34+ and AC133+ Progenitor Cells Ex-Vivo Expanded in the Presence of Tetraethylenepentamine: Reproducibility among Cord Blood Units. Blood 2004;104(suppl):405(abstract).

Hagglund H, Ringden O, Agren B, Wennberg L, Remberger M et al.

- Intraosseous compared to intravenous infusion of allogeneic bone marrow. Bone Marrow Transplant. 1998;21:331-335.
- Hock H, Hamblen MJ, Rooke HM, Schindler JW, Saleque S et al. Gfi-1 restricts proliferation and preserves functional integrity of hematopoeitic stem cells. Nature 2004;431:1002-1007.
- Ikeda Y, Fukuda N, Wada M, Matsumoto T, Satomi A et al. Development of angiogenic cell and gene therapy by transplantation of umbilical cord blood with vascular endothelial growth factor gene. Hypertens Res. 2004;27:119-128.
- Jaroscak J, Goltry K, Smith A, Waters-Pick B, Martin PL et al. Augmentation of umbilical cord blood (UCB) transplantation with ex-vivo expanded UCB cells: results of a phase I trial using the AastromReplicell System. Blood 2003;101:5061-5067.
- Kedereit S, Mohammad SF, Miller RE, Woods, KD. Listrom, CD et al. Reduced NFATI protein expression in human umbilical cord blood T lymphocytes. Blood 1999;94:3101-3107.
- Kern S, Eichler H, Stoeve J, Klüter H, Bieback K. Comparative analysis of mesenchymal stem cells from bone marrow, umbilical cord blood, or adipose tissue. Stem Cells 2006 May;24(5):1294-301.
- Kohler T, Plettig R, Wetzstein W, Scellshaffer B, Ordemann R et al. Defining optimum conditions for the ex vivo expansion of human umbilical cord blood cells. Influences of progenitor enrichment, interference with feeder layers, early-acting cytokines and agitation of culture vessels. Stem Cells 1999;17:19-24.
- Kohn DB, Hershfield MS, Carbonaro D, Shigeoka A, Brooks J et al. T lymphocytes with normal ADA gene accumulate after transplantation of transduced autologous umbilical cord blood CD34+ cells in ADAdeficient SCID neonates. Nat Med. 1998;4:775-780.
- Laughlin MJ, Eapen M, Rubinstein P, Wagner JE, Zhang MJ et al. Outcome after transplantation of cord blood or bone marrow from unrelated donors in adults with leukemia. N Engl J Med. 2004;351:2265-2275.
- Laughlin MJ, Barker J, Bambach B, Koc ON, Rizzieri DA et al. Hematopoeitic engraftment and survival in adult recipients of umbilical cord blood from unrelated donors. N Engl J Med. 2001;344;1815-1822.
- Laughlin MJ, Eapen M, Rubinstein P, Wagner JE, Zhang MJ et al. Outcome after transplantation of cord blood or bone marrow from unrelated donors in adults with leukemia. N Engl J Med. 2004;351:2265-2275.
- Lauret E, Catelain C, Titeux M, Poirault S, Dando JS et al. Membranebound delta-4 notch ligand reduces the proliferative activity of primitive human hematopoeitic CD34+CD38low cell while maintaining their LTC-IC potential. Leukemia 2004;4:788-897.
- Lauret M, Katayama N, Hoshino N, Nishikawa H, Sakano S et al. The soluble Notch ligand, Jagged-1, inhibits proliferation of macrophage progenitors. Int J Hematol. 2002;3:269-276.
- Le Blanc K, Samuelsson H, Gustafsson B, Remberger M, Sundberg B et al. Transplantation of mesenchymal stem cells to enhance engraftment of hematopoietic stem cells. Leukemia 2007 Aug;21(8):1733-8.
- Lee OK, Kuo TK, Chen WM, Lee KD, Hsieh SL et al. Isolation of multipotent mesenchymal stem cells from umbilical cord blood. Blood 2004 Mar 1;103(5):1669-75.
- Leung W, Ramirez M, Mukherjee G, Perlman EJ, Civin CL. Comparisons of alloreactive potential of clinical hematopoietic grafts. Transplantation 1999;68:628-635.
- Lewis I, Vefaillie CM. Multy-lineage expansion potential of primitive hematopoietic progenitors. Superiority of umbilical Cord blood compared

- to mobilized peripheral blood. Exp Hematol. 2000;28:1087-950.
- Li CD, Zhang WY, Li HL, Jiang XX, Zhang Y et al. Mesenchymal stem cells derived from human placenta suppress allogeneic umbilical cord blood lymphocyte proliferation. Cell Res. 2005 Jul;15(7):539-47.
- Liu E, Law HK, Lau YL. Tolerance associated with cord blood transplantation may depend on the state of host dendritic cells. Br J Haematol. 2004;126:517-526.
- Locatelli F, Rocha V, Chastang C, Arcese W, Michel G et al. Factors associated with outcome after cord blood transplantation in children with acute leukemia. Blood 1999;93:3662-3671.
- Locatelli F, Rocha V, Reed W, Bernaudin F, Ertem M et al. Related umbilical cord blood transplant in patient with thalassemia and sickle cell disease. Blood 2003;101:2137-2143.
- Long GD, Laughlin M, Madan B, Kurtzberg J, Gasparetto C et al. Unrelated umbilical cord blood transplantation in adult patients. Biol Blood Marrow. 2003;9:772-780.
- Ma N, Ladilov Y, Kaminski A, Piechaczek C, Choi YH et al. Umbilical cord blood cell transplantation for myocardial regeneration. Transplant Proc. 2006 Apr;38(3):771-3.
- Mayani H, Landsrop PM. Biology of human umbilical cord blood-derived hematopoietic stem/progenitor cells. Stem Cells 1998;16:153-165.
- Magro E, Regidor C, Cabrera R, Sanjuán I, Forès R et al. Fernandez MN. Early hematopoietic recovery after single unit unrelated cord blood transplantation in adults supported by co-infusion of mobilized stem cells from a third party donor. Haematologica 2006 May;91(5):640-8.
- McNiece I, Kubegov D, Kerzic P, Shapal EJ, Gross S. Increased expansion and differentiation of cord blood products using a two-step expansion culture. Exp Hematol. 2000;41:1567-1576.
- McNiece I, Harrington J, Turney J, Kellner J, Shpall EJ. Ex vivo expansion of cord blood mononuclear cells on mesenchymal stem cells. Cytotherapy 2004;6(4):311-7.
- Michel G, Rocha V, Chevret S, Arcese W, Chan KW, et al. Unrelated cord blood donor transplantation for children with AML a Eurocord Group analysis. Blood 2003;102:4290-4297.
- Miyakoshi S, Yuji K, Kami M, Kusumi E, Kishi Y et al. Successful engraftment after reduced-intensity umbilical cord blood transplantation for adult patients with advanced hematological disease. Clin Cancer Res. 2004;10:3586-3592.
- Milhem M, Mahmud N, Lavelle D, Araki H, DeSimone J et al. Modification of hematopoietic stem cell fate by 5aza 2' deoxycytide and trichostatin A. Blood 2004;103:4102-4110.
- Nauta AJ, Kruisselbrink AB, Lurvink E, Mulder A, Claas FH et al. Enhanced engraftment of umbilical cord blood-derived stem cells in NOD/SCID mice by cotransplantation of a second unrelated cord blood unit. Exp Hematol. 2005 Oct;33(10):1249-56.
- Newsome PN, Johannessen I, Boyle S, Dalakas E, McAulay KA et al. Human cord blood-derived cells can differentiate into hepatocytes in the mouse liver with no evidence of cellular fusion. Gastroenterology 2003;124:1891-1900.
- Nitsche A, Zhang M, Clauss T, Siegert W, Brune K et al. Cytokine profiles of cord and adult blood leukocytes: differences in expression are due to differences in expression and activation of transcription factors. BMC Immunol. 2007 Aug 31;8:18.
- Nolta JA, Thiemann FT, Arakawa-Hoyt J, Dao MA, Barsky LW et al. The AFT024 stromal cell line supports long term ex-vivo maintenance of engrafting multipotent human hematopoeitic progenitors. Leukemia 2002;16:352-361.

Pecora AL, Stiff P, Jennis A, Goldberg S, Rosenbluth R et al. Prompt and durable engraftment in two adult patients with high risk chronic myelogenous leukemia (CML) using ex-vivo expanded and unmanipulated unrelated umbilical cord blood. Bone marrow Transplant. 2000;25:797-

Peled T, Landau E, Mandel J, Glukhman E, Goudsmid NR et al. Linear polyamine copper chelator tetraethylenepentamine augments long-term ex vivo expansion of cord blood-derived CD34+ cells and increases their engraftment potential in NOD/SCID mice. Exp Hematol. 2004;32:547-

Peled T, Rubinstein P, Kurtzberg J. TEPA Augments Ex-Vivo and In-Vivo Potential of Cord Blood Derieved CD34+ Cells: From Basic Science to Clinical Trials. Blood 2004:111(suppl).3581(abstract).

Peled T, Mandel J, Goudsmid RN, Landor C, N Hasson, D Harati et al. Pre-Clinical Development of Cord Blood-Derived Progenitor Cell Graft Expanded Ex Vivo with cytokines and Polyamine Copper Chelator Tetraethylenepentamine. Cytotherapy 2004;6:344-355.

Pesce M, Orlandi A, Iachininoto MG, Straino S, Torella AR et al. Myoendothelial Differentiation of Human Umbilical Cord Blood-Derived Stem Cells in Ischemic Limb Tissues. Circ. Res. 2003;93:e51–62.

Piacibello W, Gammaitoni L, Bruno S, Gunetti M, Fagioli F et al. Neagtive influence of IL3 on the expansion of human cord blood in vivo long-term repopulating stem cells. J Hematother Stem Cell Res. 2000;9:945-956.

Pick M, Nagler A, Grisaru D, Eldor A, Deutsch V. Expansion of megakaryocyte progenitors from human umbilical cord using a new twostep separation procedure. Br J Haematol. 1998;103:639-650.

Risdon G, Gaddy J, Stehman FB, Broxmeyer HE. Proliferative and cytotoxic responses of human cord blood T lymphocytes following allogeneic stimulation. Cell Immunol. 1994;154:14-24.

Risdon G, Gaddy J, Horie M, Broxmeyer HE. Alloantigen priming induces a state of unresponsiveness in human cord blood T cells. Proc Natl Acad Sience USA. 1995;92:2413-2417.

Robinson SN, Ng J, Niu T, Yang H, McMannis JD et al. Superior ex vivo cord blood expansion following co-culture with bone marrow-derived mesenchymal stem cells. Bone Marrow Transplant. 2006 Feb;37(4):359-

Rocha V, Labopin M, Sanz G, Arcese W, Scellshwerdtfeger R et al. Transplants of umbilical-cord blood or bone marrow from unrelated donors in adults with acute leukemia. N Engl J Med. 2004;351:2276-

Rocha V, Wagner JE, Sobocinski KA, Klein JP, Zhang MJ et al. Graft-versus-host disease in children who have received a cord-blood or bone marrow transplant from HLA-identical sibling. N Engl J Med. 2000;342:1846-1854.

Rocha V, Gluckman E. Outcomes of transplantation in children with acute leukaemia. Lancet 2007 Jun 9;369(9577):1906-8.

Roncarolo MG, Bigler M, Martino S, Ciuti E, Tovo PA et al. Immune functions of cord blood cells before and after transplantation. J Hematolther. 1996;5:157-160.

Rubinstein P, Stevens CE. Placental blood for bone marrow replacement: the New York Blood Center's program and clinical results. Baillieres Best Pract Res Clin Haematol. 2000;13:565-58.

Secco M, Zucconi E, Vieira NM, Fogaça LL, Cerqueira A et al. Multipotent stem cells from umbilical cord: cord is richer than blood! Stem Cells 2008 Jan;26(1):146-506.

Shpall EJ, Quinones R, Giller R, Zeng C, Baron AE et al. Transplantation of ex vivo expanded cord blood. Biol of Blood and Marrow Trans. 2002;8:368-376.

Shpall EJ, De Lima M, Chan K, Champlin R, Gee A et al. A Phase I/II study of ex-vivo expanded cord blood for leukemia and Lymphoma. Cytotherapy 2005;7 (suppl 1):221 (abstract).

Shpall; EJ, De Lima M, Chan K, Champlin A, Gee P et al. Transplantation of cord blood expanded ex vivo with copper chelator. Blood 2004;104(s uppl):982(abstract).

Sorg RV, Kogler G, Wernet P. Functional competence of dendritic cells in human umbilical cord blood. Bone Maroow Transplant. 1998;22(Suppl 1):S52-54.

Staba SL, Escolar ML, Poe M, Kim Y, Martin PL et al. Cord-Blood transplants from unrelated donors in patients with Hurler's syndrome. N Engl J Med. 2004;350:1960-1969.

Stein J, Zion Garty B, Dror Y, Fenig E, Zeigler M et al. Successful treatment of Wolman disease by unrelated umbilical cord blood transplantation. Eur J Pediatr. 2007 Jul;166(7):663-6.

Takahashi S, Iseki T, Ooi J, Tomonari A, Takasugi K et al. Single-institute comparative analysis of unrelated bone marrow transplantation and cord blood transplantation for adult patients with hematologic malignancies. Blood 2004 Dec 1;104(12):3813-20.

Takahashi S, Ooi J, Tomonari A, Konuma T, Tsukada N, et al. Comparative single-institute analysis of cord blood transplantation from unrelated donors with bone marrow or peripheral blood stemcell transplants from related donors in adult patients with hematologic malignancies after myeloablative conditioning regimen. Blood 2007 Feb 1;109(3):1322-30.

Vendrame M. Cassady J. Newcomb J. Butler T. Pennypacker KR et al. Infusion of human umbilical cord cells in a rat model of stroke dose dependently rescues behavioral deficits and reduces infarct volume. Stroke 2004;35:2390-2395.

Verneris MR, Brunstein CG, DeFor T, Barker J, Weisdorf DJ et al. Risk of relapse after umbilical cord blood transplantation in patients with acute leukemia: marked reduction in recipients of two units. ASH Annual Meeting Abstracts 2005;106:305.

Wagner JE, Barker JN, DeFor TE, Baker KS, Blazar BR et al. Transplantation of unrelated donor umbilical cord blood in 102 patients with malignant and nonmalignant diseases: influence of CD34 cell dose and HLA disparity on treatment-related mortality and survival. Blood 2002;100:1611-1618.

Wang J, Kimura T, Asada R, Harada S, Yokota S et al. SCID-repopulating cell activity of human cord blood-derived CD34- cells assured by intrabone marrow injection. Blood 2003;101:2924-2931.

Yahata T, Ando K, Sato T, Miyatake H, Nakamura Y et al. A highly sensitive strategy for SCID-repopulating cell assay by direct injection of primitive human hematopoietic cells into NOD/SCID mice bone marrow. Blood 2003;101:2905-2913.

Yuji K, Miyakoshi S, Kato D, Miura Y, Myojo T et al. Reduced-Intensity Unrelated Cord Blood Transplantation for patients with Advanced Malignant Lymphoma. Biol Blood Marrow Trans. 2005;11:314-318.

Zhang Y, Li C, Jiang X, Zhang S, Wu Y et al. Human placental-derived mesenchymal progenitor cells support expansion of long-term cultureinitiating cells from cord blood CD34+ cells. Exp Hematol. 2004;32:657-664.

© The Authors. This article is provided under the following license: Creative Commons Attribution-Noncommercial 2.0 Germany, http://creativecommons.org/licenses/by-nc/2.0/de/deed.en

Пуповинная кровь: от побочного продукта родов до ценного источника жизнеспасающего лечения

Гэл Гольдштейн, Амос Торен, Арнон Наглер

Расширенное резюме

Обзор посвящен вопросам трансплантации гемопоэтических стволовых клеток из пуповинной крови (ГСК ПК), который ранее применялся в детской практике. Кратко перечислены процедуры сбора ГСК ПК во время родов, а также рутинные тесты оценки их качества (НLА-типирование, проверка инфекционных агентов). Сейчас в мире около 250000 доз ГСК ПК хранятся в 35 банках 21 страны. Этические проблемы с применением клеток ПК могут возникать при их длительном хранении. Указывается на противоречия, связанные с развитием частных банков пуповинной крови (по оценкам, в них хранятся ок.600000 доз ПК), ввиду неопределенности сроков гарантированного хранения стволовых клеток для возможной трансплантации. Свойства ПК как источника ГСК ограничены небольшим объемом образца и малым числом ГСК, обладающих высокой пролиферативной активностью, при меньшем содержании Т-клеток и их большей иммунологической толерантностью. Это дает возможность проводить пересадки, с меньшими ограничениями по HLA-совместимости, при меньшем риске отторжения и более низкой частоте РТПХ у больных.

Авторы обобщают клинический опыт ТГСК ПК в различных центрах, где показана высокая эффективность этого метода при более длительных сроках восстановления гемопоэза, чем трансплантации костного мозга. Минимально допустимой дозой ГСК ПК считается 1,5-2,5Х107 миелокариоцитов на 1 кг массы тела больного. Описываются основные области применения ГСК ПК (родственная или неродственная трансплантация у детей при неопухолевых и злокачественных и заболеваниях). Подчеркивается нехватка сравнительных данных об эффективности ГСК из пуповинной крови и костного мозга.

Особое внимание уделяется ТГСК ПК при неопухолевых заболеваниях с аплазией костного мозга, где риск неприживления оказался недопустимо высоким. Описаны хорошие результаты ТГСК ПК при гемоглобинопатиях, мукополисахаридозах. При лечении взрослых больных посредством ТГСК ПК предлагаются немиелоаблативные режимы кондиционирования, хотя эффективность такого подхода пока неясна. Обсуждается возможность одновременной трансплантации двух и более доз ГСК от разных доноров, включая дозу ПК. Дискутируется вопрос о внутрикостном введении ГСК ПК, разрабатываются методы культивирования ГСК ПК в культуре, хотя темпы их размножения этих клеток пока недостаточны, а их способность к дифференцировке мало изучена. В качестве добавочного стимула предложено введение мезенхимных стволовых клеток совместно с ГСК ПК. В заключение описывается использование нормальных ГСК ПК для коррекции генетических дефектов у детей, а также их плюрипотентность для репарации дефектов других тканей (например, миокарда или сосудов).

Ключевые слова: пуповинная кровь, стволовые кроветворные клетки, заготовка, хранение, трансплантация, обзор